

A. Salakka and K. Wähälä*

Laboratory of Organic Chemistry, Department of Chemistry, PO Box 55, FIN-00014, University of Helsinki, Finland. E-mail: Kristiina.Wahala@helsinki.fi

Received (in Cambridge, UK) 21st June 1999, Accepted 30th July 1999

Reduction of hydroxy and/or methoxy-substituted isoflavones using LiAlH_4 in refluxing THF provides an easy access to a number of α -methyldeoxybenzoins, possible metabolites of phytoestrogens in man. The synthesis of *O*-demethylangolensin **2b**, 6'-hydroxyangolensin **2c**, angolensin **2d**, 1-(2,4-dihydroxyphenyl)-2-phenylpropan-1-one **2e**, 1-(2-hydroxy-4-methoxyphenyl)-2-(4-hydroxyphenyl)propan-1-one **2f**, 4'-*O*-methylangolensin **2g**, 1-(2-hydroxy-4-methoxyphenyl)-2-phenylpropan-1-one **2h**, and 1-(2-hydroxyphenyl)-2-phenylpropan-1-one **2i** is described.

Recently, it has been established that α -methyldeoxybenzoins are likely metabolites of naturally occurring isoflavones such as genistein **1a**, daidzein **1b**, biochanin A **1c** and formononetin **1d**. The α -methyldeoxybenzoins as such are rare in the plant kingdom. The sole known example is angolensin **2d** which occurs in the heartwood of *Pterocarpus angolensis*, in *P. indicana* and in *Afromosia* species.¹⁻⁵ Angolensin methyl^{1,2} and cadinyl (sesquiterpenoid) ethers^{1,2} are also known to occur naturally.

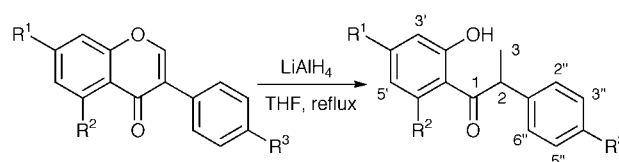
Interest in the metabolism of the dietary isoflavones in man is now rapidly increasing. It is now known that formononetin **1d** is first converted to daidzein **1b** and thence to *O*-demethylangolensin (ODMA, **2b**),⁶ a member of the α -methyldeoxybenzoin group of compounds. ODMA **2b** was first identified from the urine and plasma of sheep⁷⁻⁹ and cattle.^{6,9} Later it was isolated from the urine of chimpanzees¹⁰ and humans,¹¹⁻¹³ human plasma,^{14,15} human faeces,^{16,17} and from cow milk.¹⁸ Angolensin **2d**, the methylated analog of ODMA, has been found only in the urine of sheep.¹⁹ The tentative identification of a new α -methyldeoxybenzoin, 6'-hydroxy-*O*-demethylangolensin **2a**, in human urine was recently announced.^{20,21} We have since confirmed this by comparison with an authentic synthetic sample.²²

Known syntheses of α -methyldeoxybenzoins mostly encompass numerous steps giving poor overall yields, as, for example, in the synthesis of 4'-hydroxy-2',4'-dimethoxy- α -methyldeoxybenzoin by an 8-step route involving the photolysis of an α -hydroxydihydrochalcone.⁵ Similarly, there is a multistep route to angolensin and ODMA by way of a Friedel-Crafts reaction of 1,3-dimethoxybenzene and 2-methoxypropionic acid.^{11,23} More recently, α -methyldeoxybenzoins have been prepared by a base-catalysed C-methylation of deoxybenzoins, giving angolensin and its methylated analogs.²⁴ A free OH at C-2' is compatible whereas a 4'-OH is methylated. Our experience is that 2',4'-dihydroxydeoxybenzoins give complex mixtures of products, also by 4'- and α,α -dialkylation, with various procedures under basic conditions.²⁵

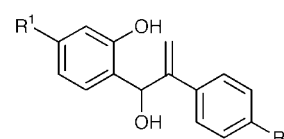
O-Methylated and -benzylated analogs of angolensin have been prepared directly from angolensin by methylation^{1,26,27} or benzylation.¹

Certain reductions of isoflavones are reported to furnish α -methyldeoxybenzoins. Catalytic hydrogenation of the unsubstituted isoflavone in basic medium leads mainly to 2'-hydroxy- α -methyldeoxybenzoin presumably *via* the ring-opened α -methylene derivative.²⁸ Also, Birch reduction using lithium in liquid ammonia converts protected isoflavones to the corresponding α -methyldeoxybenzoins.²⁹ Some reductions of protected isoflavones with LiAlH_4 giving ring-opened ketones and further α -methyldeoxybenzoins as minor products have

also been reported.³⁰ However, there are no mentions of hydride reductions of unprotected hydroxy-substituted isoflavones in the literature. The co-occurrence of α -methyldeoxybenzoins with various isoflavonoid derivatives, and their origin as reduced metabolites of the latter, prompted us to study the synthesis of α -methyldeoxybenzoins directly from isoflavone starting materials by reduction. This would be highly expedient as we already have an easy and general access to a number of isoflavones by a one-pot reaction.^{31,32} Following our preliminary report of the first synthesis of 6'-hydroxy-*O*-demethylangolensin **2a**, by reduction of genistein with LiAlH_4 in refluxing THF,³³ we now describe an improved synthetic pathway and spectroscopic details for angolensin **2d**, *O*-demethylangolensin **2b** and for seven additional α -methyldeoxybenzoins, many of them new (Table 1) (see Scheme 1).



1a $\text{R}^1 = \text{R}^2 = \text{R}^3 = \text{OH}$	2a $\text{R}^1 = \text{R}^2 = \text{R}^3 = \text{OH}$
1b $\text{R}^1 = \text{R}^3 = \text{OH}, \text{R}^2 = \text{H}$	2b $\text{R}^1 = \text{R}^3 = \text{OH}, \text{R}^2 = \text{H}$
1c $\text{R}^1 = \text{R}^2 = \text{OH}, \text{R}^3 = \text{OCH}_3$	2c $\text{R}^1 = \text{R}^2 = \text{OH}, \text{R}^3 = \text{OCH}_3$
1d $\text{R}^1 = \text{OH}, \text{R}^2 = \text{H}, \text{R}^3 = \text{OCH}_3$	2d $\text{R}^1 = \text{OH}, \text{R}^2 = \text{H}, \text{R}^3 = \text{OCH}_3$
1e $\text{R}^1 = \text{OH}, \text{R}^2 = \text{R}^3 = \text{H}$	2e $\text{R}^1 = \text{OH}, \text{R}^2 = \text{R}^3 = \text{H}$
1f $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{H}, \text{R}^3 = \text{OH}$	2f $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{H}, \text{R}^3 = \text{OH}$
1g $\text{R}^1 = \text{R}^3 = \text{OCH}_3, \text{R}^2 = \text{H}$	2g $\text{R}^1 = \text{R}^3 = \text{OCH}_3, \text{R}^2 = \text{H}$
1h $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{R}^3 = \text{H}$	2h $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{R}^3 = \text{H}$
1i $\text{R}^1 = \text{R}^2 = \text{R}^3 = \text{H}$	2i $\text{R}^1 = \text{R}^2 = \text{R}^3 = \text{H}$



3a $\text{R}^1 = \text{R}^2 = \text{OCH}_3$
3b $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{H}$
3c $\text{R}^1 = \text{R}^2 = \text{H}$
3d $\text{R}^1 = \text{OCH}_3, \text{R}^2 = \text{OH}$

Scheme 1 Reduction of isoflavones with LiAlH_4 .

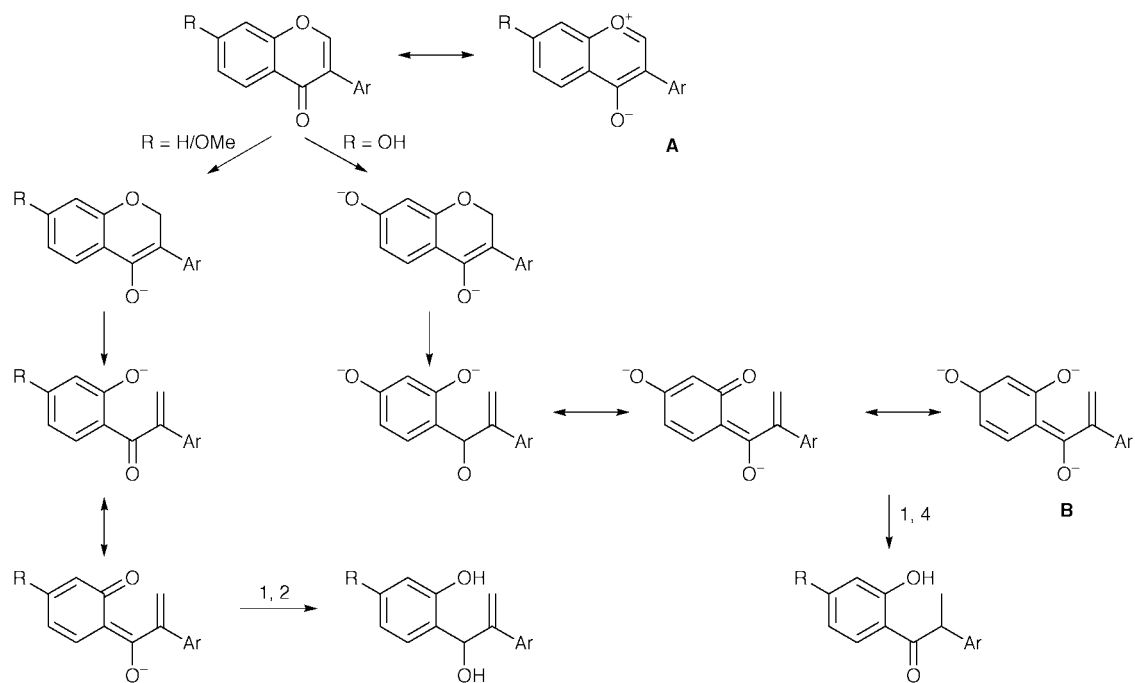


Table 1 Reduction of isoflavones with LiAlH₄ in refluxing THF

Starting material	Amount of LiAlH ₄ (mol equiv.)	Reaction time (t/h)	Products	Yield (%)
1a	5.5	2.5	2a ³³	66
1b	5.5	24	2b	42
1c	4.3	1	2c ^a	70
1d	3.3	2	2d	42
1e	3.3	1.5	2e ^a	60
1f	3.3	1.5	2f ^a	34
			3d ^a	17
1g	2.5	0.25	3a ^a	50
			2g	27
1h	2.5	0.25	3b ^a	50
			2h	29
1i	2.5	0.25	3c ^a	48
			2i	27

^a New compound.

Results and discussion

As seen in the Table, the amount of reagent used was adjusted to take into account the number of free OH groups. It is also clearly seen how an increasing number of free OH groups requires a longer reaction time (in refluxing THF), presumably due to a combination of electronic and solubility effects.

The α -methyldeoxybenzoin is formed without complications except when there is no hydroxy substituent at the isoflavone 7-position. In such cases prop-2-en-1-ols are the main products. For example, the reduction of 4',7-dimethoxyisoflavone **1g** gives 1-(2-hydroxy-4-methoxyphenyl)-2-(4-methoxyphenyl)prop-2-en-1-ol **3a** and 4'-*O*-methylangolensin **2g** in a ratio of 2 to 1. Contrary to some earlier proposals,²⁸ we assume that the initial step in these reductions is a 1,4-addition to the enone system, driven by the 4-oxypyrylium contributor (A) to the isoflavone structure. This is then followed by ring opening to a prop-2-en-1-one, in turn reduced by another 1,4-addition to give the deoxybenzoin enolates, or by a 1,2-addition to give the propenols (Scheme 2). We suggest that the effect of the 7-hydroxy substituent is to provide a further resonance contributor (B) where the propenone carbonyl is lacking, thus favouring conjugate hydride attack at the propene terminus.

As the situation is somewhat different with the 7-H or 7-MeO analogs 1,2-attack at the propenone carbonyl predominates.

Experimental

Mps were determined in open capillary tubes with an Electrothermal apparatus and are uncorrected. IR spectra were recorded from KBr discs on a FTIR Biorad FTS-7 instrument. UV spectra were recorded in 94% ethanol solution with a CARY 5E UV-VIS-NIR spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Varian GEMINI-200 FT NMR spectrometer using the standard ¹H/¹³C dual probe (chemical shifts in ppm). *J*-Values are given in Hz. LR and HR mass spectra were obtained with a JEOL JMS SX102 mass spectrometer operating at 70 eV. TLC was conducted on Merck silica gel 60 F₂₅₄ plates, and Merck silica gel 60 (0.040–0.063 mm, 230–400 mesh) was used for flash chromatography. Tetrahydrofuran (THF) was dried by distilling over CaH₂ before use.

General procedure

In Table 1 there are collected the starting isoflavone **1a–i**, the amount of reducing agent used and the reaction time needed for the completion of reduction is judged by TLC monitoring. A solution of an isoflavone **1** (0.2 g) in THF (10 cm³) was added over 0.5 h to a stirred slurry of lithium aluminium hydride (2.5–5.5 mol equiv., see Table 1) in refluxing THF (5 cm³). After further refluxing (see Table 1), the reaction mixture was cooled and poured into saturated aq. NH₄Cl at 0 °C. The mixture was neutralized with 2 M HCl and extracted with EtOAc. The extract was washed with water, dried over Na₂SO₄ and evaporated. The residue was purified by flash chromatography.

O-Demethylangolensin **2b**, 1-(2,4-dihydroxyphenyl)-2-(4-hydroxyphenyl)propan-1-one

Flash chromatography eluent: *n*-hexane–acetone (1:1). White crystals, mp 101 °C (from *n*-hexane–diethyl ether) (lit.,³⁴ 103 °C); λ_{\max} (EtOH)/nm 214 (ϵ /dm³ mol⁻¹ cm⁻¹ 21 700), 278 (15 300) and 318 (9600); ν_{\max} /cm⁻¹ 3303 (OH) and 1631 (CO); δ_{H} ([²H₆]acetone) 1.43 (3H, d, *J* 6.8, 3-H₃), 4.76 (1H, q, *J* 6.8, 2-H), 6.30 (1H, d, *J* 2.4, 3'-H), 6.37 (1H, dd, *J* 2.4 and 8.6, 5'-H), 6.78 (2H, d, *J* 8.6, 3''- and 5''-H), 7.20 (2H, d, *J* 8.5, 2''-H and 6''-H) and 7.89 (1H, d, *J* 8.8, 6'-H); δ_{C} ([²H₆]acetone) 18.1

(C-3), 44.8 (C-2), 102.3 (C-3'), 107.3 (C-5'), 111.6 (C-1'), 115.1 (C-3'' and -5''), 128.1 (C-2'' and -6''), 132.3 (C-1''), 132.7 (C-6'), 155.8 (C-4''), 164.0 (C-2'), 165.6 (C-4') and 204.8 (C-1); *m/z* 258 (M⁺, 14%), 138 (8), 137 (100) and 121 (17) (Found: M⁺, 258.0894. C₁₅H₁₄O₄ requires *M*, 258.0892).

6'-Hydroxyangolensin 2c, 2-(4-methoxyphenyl)-1-(2,4,6-tri-hydroxyphenyl)propan-1-one

Flash chromatography eluent: *n*-hexane–acetone (1:1). White crystals, mp 107 °C (from benzene); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 225 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 11 500) and 294 (11 100); $\nu_{\max}/\text{cm}^{-1}$ 3347 (OH) and 1630 (CO); $\delta_{\text{H}}([\text{C}_6\text{H}_6]\text{acetone})$ 1.40 (3H, d, *J* 7.0, 3-H₃), 3.73 (3H, s, OCH₃), 5.27 (1H, q, *J* 6.9, 2-H), 5.89 (2H, s, 3'- and 5'-H), 6.8 (2H, d, *J* 8.8, 3''-H and 5''-H) and 7.23 (2H, d, *J* 8.8, 2''-H and 6''H); $\delta_{\text{C}}([\text{C}_6\text{H}_6]\text{acetone})$ 19.9 (C-3), 49.4 (C-2), 55.3 (OCH₃), 96.0 (C-3' and -5'), 104.8 (C-1'), 114.4 (C-3'' and -5''), 130.0 (C-2'' and -6''), 135.3 (C-1''), 159.3 (C-4''), 165.3 (C-2', -4' and -6') and 207.1 (C-1); *m/z* 288 (M⁺, 18%), 270 (5), 167 (6), 153 (100) and 135 (45) (Found: M⁺, 288.1000. C₁₆H₁₆O₅ requires *M*, 288.0998).

Angolensin 2d, 1-(2,4-dihydroxyphenyl)-2-(4-methoxyphenyl)-propan-1-one

Flash chromatography eluent: CH₂Cl₂–EtOAc (7:1). Pale yellowish crystals, mp 84 °C (from benzene) (lit.,²⁴ 86–87 °C); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 215 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 37 400), 280 (26 800) and 319 (17 000); $\nu_{\max}/\text{cm}^{-1}$ 3353 (OH) and 1630 (CO); $\delta_{\text{H}}(\text{CDCl}_3)$ 1.49 (3H, d, *J* 7.0, 3-H₃), 3.75 (3H, s, 4'-OMe), 4.57 (1H, q, *J* 6.8, 2-H), 6.27 (1H, d, *J* 2.5, 3'-H), 6.32 (1H, dd, *J* 2.6 and 8.8, 5'-H), 6.84 (2H, d, *J* 8.6, 3''- and 5''-H), 7.20 (2H, d, *J* 8.7, 2''- and 6''-H), 7.68 (1H, d, *J* 8.8, 6'-H) and 12.00 (1H, br s, OH); $\delta_{\text{C}}(\text{CDCl}_3)$ 19.2 (C-3), 45.9 (C-2), 55.3 (OMe), 103.5 (C-3'), 108.1 (C-5'), 112.8 (C-1'), 114.4 (C-3'' and -5''), 128.6 (C-2'' and -6''), 132.8 (C-6'), 133.5 (C-1''), 158.4 (C-4''), 162.8 (C-2'), 165.6 (C-4') and 205.4 (C-1); *m/z* 272 (M⁺, 36%), 151 (5), 149 (5), 138 (17), 137 (100), 136 (15), 135 (60), 105 (5) and 91 (5) (Found: M⁺, 272.1047. C₁₆H₁₆O₄ requires *M*, 272.1048).

1-(2,4-Dihydroxyphenyl)-2-phenylpropan-1-one 2e

Flash chromatography eluent: CH₂Cl₂–EtOAc (7:2). White crystals, mp 137 °C (from *n*-hexane–Et₂O); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 213 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 22 200), 282 (15 300), 234 (8650) and 317 (9540); $\nu_{\max}/\text{cm}^{-1}$ 3357 (OH) and 1631 (CO); $\delta_{\text{H}}([\text{C}_6\text{H}_6]\text{acetone})$ 1.48 (3H, d, *J* 6.8, 3-H₃), 4.88 (1H, q, *J* 6.6, 2-H), 6.30 (1H, d, *J* 2.2, 3'-H), 6.36 (1H, dd, *J* 2.4 and 8.8, 5'-H), 7.16–7.43 (5H, m, Ar-H) and 7.90 (1H, d, *J* 9.0, 6'-H); $\delta_{\text{C}}([\text{C}_6\text{H}_6]\text{acetone})$ 19.5 (C-3), 47.1 (C-2), 103.7 (C-3'), 108.8 (C-5'), 113.1 (C-1'), 127.8 (C-4''), 128.5 (C-2'' and -6''), 129.8 (C-3'' and -5''), 134.1 (C-6'), 143.1 (C-1''), 165.5 (C-2'), 167.1 (C-4') and 205.8 (C-1); *m/z* 242 (M⁺, 12%), 138 (10), 137 (100), 105 (13), 77 (5) and 58 (46) (Found: M⁺, 242.0944. C₁₅H₁₄O₃ requires *M*, 242.0943).

1-(2-Hydroxy-4-methoxyphenyl)-2-(4-hydroxyphenyl)propan-1-one 2f

Flash chromatography eluent: *n*-hexane–acetone (1:1). White crystals, mp 142 °C (from *n*-hexane–diethyl ether); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 214 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 24 800), 227s (17 600), 276 (18 100) and 320 (10 400); $\nu_{\max}/\text{cm}^{-1}$ 3465 (OH) and 1629 (CO); $\delta_{\text{H}}([\text{C}_6\text{H}_6]\text{-acetone})$ 1.43 (3H, d, *J* 6.8, 3-H₃), 3.83 (3H, s, OCH₃), 4.79 (1H, q, *J* 6.9, 2-H), 6.38 (1H, d, *J* 2.6, 3'-H), 6.41 (1H, dd, *J* 8.2 and 2.6, 5'-H), 6.78 (2H, d, *J* 8.6, 3''- and 5''-H), 7.20 (2H, d, *J* 8.6, 2''- and 6''-H) and 7.94 (1H, d, *J* 8.3, 6'-H); $\delta_{\text{C}}([\text{C}_6\text{H}_6]\text{acetone})$ 19.5 (C-3), 46.4 (C-2), 56.1 (OCH₃), 101.8 (C-3'), 108.1 (C-5'), 115.2 (C-1'), 116.7 (C-3'' and -5''), 129.6 (C-2'' and -6''), 133.7 (C-6'), 134.2 (C-1''), 157.5 (C-4''), 167.1 (C-2' and -4') and 207.1 (C-1); *m/z* 272 (M⁺, 18%), 152 (15), 151 (100), 137 (14), 135 (8), 121 (8) (Found: M⁺, 272.1052. C₁₆H₁₆O₄ requires *M*, 272.1048).

4'-O-Methylangolensin 2g, 1-(2-hydroxy-4-methoxyphenyl)-2-(4-methoxyphenyl)propan-1-one

Flash chromatography eluent: *n*-hexane–acetone (7:4). White crystals, mp 67 °C (from *n*-hexane–Et₂O) (lit.,²⁴ 69–70 °C); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 215 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 25 500), 227s (18 700), 276 (18 400) and 319 (11 000); $\nu_{\max}/\text{cm}^{-1}$ 3439 (OH) and 1637 (CO); $\delta_{\text{H}}(\text{CDCl}_3)$ 1.50 (3H, d, *J* 6.8, 3-H₃), 3.75 (3H, s, 4'-OCH₃), 3.78 (3H, s, 4'-OCH₃), 4.59 (1H, q, *J* 7.0, 2-H), 6.33 (1H, dd, *J* 8.8 and 2.5, 5'-H), 6.38 (1H, d, *J* 2.3, 3'-H), 6.84 (2H, d, *J* 8.8, 3''- and 5''-H), 7.21 (2H, d, *J* 8.8, 2''- and 6''-H) and 7.69 (1H, d, *J* 8.8, 6'-H); $\delta_{\text{C}}(\text{CDCl}_3)$ 19.2 (C-3), 46.0 (C-2), 55.2 (OCH₃), 55.5 (OCH₃), 101.0 (C-3'), 107.6 (C-5'), 112.7 (C-1'), 114.3 (C-3'' and -5''), 128.5 (C-2'' and -6''), 131.9 (C-6'), 133.5 (C-1''), 158.5 (C-4''), 165.8 (C-2'), 166.1 (C-4') and 204.9 (C-1); *m/z* 286 (M⁺, 30%), 284 (15), 269 (10), 151 (100), and 135 (40) (Found: M⁺, 286.1202. C₁₇H₁₈O₄ requires *M*, 286.1205).

1-(2-Hydroxy-4-methoxyphenyl)-2-phenylpropan-1-one 2h

Flash chromatography eluent: *n*-hexane–acetone (7:4). White crystals, mp 109 °C (from *n*-hexane) (lit.,²⁹ 108 °C); $\lambda_{\max}(\text{EtOH})/\text{nm}$ 213 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 18 100), 280 (13 300), 231 (7770) and 318 (7570); $\nu_{\max}/\text{cm}^{-1}$ 3428 (OH) and 1630 (CO); $\delta_{\text{H}}(\text{CDCl}_3)$ 1.53 (3H, d, *J* 7.0, 3-H₃), 3.79 (3H, s, 4'-OCH₃), 4.64 (1H, q, *J* 6.8, 2-H), 6.32 (1H, dd, *J* 2.6 and 8.8, 5'-H), 6.39 (1H, d, *J* 2.4, 3'-H), 7.19–7.32 (5H, m, Ar-H) and 7.69 (1H, d, *J* 8.8, 6'-H); $\delta_{\text{C}}(\text{CDCl}_3)$ 19.2 (C-3), 47.0 (C-2), 55.6 (OMe), 101.1 (C-3'), 107.7 (C-5'), 112.8 (C-1'), 127.1 (C-4'), 127.6 (C-2'' and -6''), 129.1 (C-3'' and -5''), 132.1 (C-6'), 141.6 (C-1''), 165.9 (C-2'), 166.2 (C-4') and 204.7 (C-1); *m/z* 256 (M⁺, 7%), 162 (5), 152 (10), 151 (100), 117 (10) and 58 (40) (Found: M⁺, 256.1090. C₁₆H₁₆O₃ requires *M*, 256.1099).

1-(2-Hydroxyphenyl)-2-phenylpropan-1-one 2i

Flash chromatography eluent: *n*-hexane–acetone (7:4). Colorless oil; $\lambda_{\max}(\text{EtOH})/\text{nm}$ 213 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 19 100), 255 (9140) and 329 (4050); $\nu_{\max}/\text{cm}^{-1}$ 2978 (OH) and 1638 (CO); $\delta_{\text{H}}(\text{CDCl}_3)$ 1.54 (3H, d, *J* 7.0, 3-H₃), 4.73 (1H, q, *J* 6.6, 2-H), 6.78 (1H, t, *J* 8.3, 5'-H), 6.94 (1H, d, *J* 7.4, 3'-H), 7.19–7.42 (6H, m, Ar-H) and 7.79 (1H, d, *J* 8.0, 6'-H); $\delta_{\text{C}}(\text{CDCl}_3)$ 19.3 (C-3), 47.3 (C-2), 118.7 (C-1' and -3'), 118.9 (C-5'), 127.2 (C-4''), 127.6 (C-2'' and -6''), 129.1 (C-3'' and -5''), 130.5 (C-6'), 136.2 (C-4'), 141.2 (C-1''), 163.2 (C-2') and 206.6 (C-1); *m/z* 226 (M⁺, 11%), 194 (5), 122 (8), 121 (100), 105 (12), 77 (7) and 58 (35) (Found: M⁺, 226.0996. C₁₅H₁₄O₂ requires *M*, 226.0994).

1-(2-Hydroxy-4-methoxyphenyl)-2-(4-methoxyphenyl)prop-2-en-1-ol 3a

Flash chromatography eluent: *n*-hexane–acetone (7:4). Amorphous solid; $\lambda_{\max}(\text{EtOH})/\text{nm}$ 205 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 13 100) and 254 (5130); $\nu_{\max}/\text{cm}^{-1}$ 3350 (OH); $\delta_{\text{H}}(\text{CDCl}_3)$ 3.70 (3H, s, OCH₃), 3.75 (3H, s, OCH₃), 5.21 (1H, s, 3-H), 5.43 (1H, s, 3-H), 5.80 (1H, s, 1-H), 6.32 (1H, dd, *J* 8.4 and 2.4, 5'-H), 6.41 (1H, d, *J* 2.4, 3'-H), 6.78–6.86 (3H, m, 6'-, 3''- and 5''-H), 7.30 (2H, d, *J* 6.7, 2''- and 6''-H) and 7.97 (1H, br s, OH); $\delta_{\text{C}}(\text{CDCl}_3)$ 55.2 (2 × OCH₃), 75.4 (C-1), 102.4 (C-3), 105.8 (C-3'), 113.6 (C-5'), 113.9 (C-3'' and -5''), 117.6 (C-1'), 128.1 (C-2'' and -6''), 129.2 (C-6'), 131.0 (C-1''), 148.0 (C-2), 156.9 (C-2'), 159.2 (C-4') and 160.5 (C-4''); *m/z* 286 (M⁺, 2%), 282 (20), 269 (18), 268 (100, M⁺ – 18), 267 (80), 253 (21), 239 (10), 224 (8), 153 (17) and 134 (10) (Found: M⁺, 286.1219. C₁₇H₁₈O₄ requires *M*, 286.1176).

1-(2-Hydroxy-4-methoxyphenyl)-2-phenylprop-2-en-1-ol 3b

Flash chromatography eluent: *n*-hexane–acetone (7:4). Amorphous solid. $\lambda_{\max}(\text{EtOH})/\text{nm}$ 205 ($\epsilon/\text{dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ 27 300), 236 (10 200) and 282 (2670); $\nu_{\max}/\text{cm}^{-1}$ 3449 (OH) and 3146 (OH); $\delta_{\text{H}}(\text{CDCl}_3)$ 3.72 (3H, s, 4'-OCH₃), 5.31 (1H, s, 3-H), 5.51 (1H, s, 3-H), 5.85 (1H, s, 1-H), 6.34 (1H, dd, *J* 2.5 and 8.5,

5'-H), 6.44 (1H, d, *J* 2.6, 3'-H), 6.86 (1H, d, *J* 8.4, 6'-H) and 7.20–7.41 (5H, m, Ar-H); δ_{C} (CDCl₃) 55.8 (OMe), 75.8 (C-1), 103.0 (C-3), 106.4 (C-3'), 115.6 (C-5'), 118.3 (C-1'), 127.6 (C-2'' and -6''), 128.5 (C-4''), 129.0 (C-3'' and -5''), 129.8 (C-6'), 139.5 (C-1''), 149.5 (C-2), 157.5 (C-2') and 161.1 (C-4'); *m/z* 254 (M⁺, 5%), 239 (18), 238 (M⁺ - 18, 100), 237 (68), 223 (13), 194 (7), 165 (12), 161 (22), 117 (11) and 58 (34) (Found: M⁺, 256.1084. C₁₆H₁₆O₃ requires *M*, 256.1099).

1-(2-Hydroxyphenyl)-2-phenylprop-2-en-1-ol 3c

Flash chromatography eluent: *n*-hexane–acetone (7:4). Amorphous solid. λ_{max} (EtOH)/nm 205 ($\epsilon/\text{dm}^3 \text{mol}^{-1} \text{cm}^{-1}$ 24 500), 240 (8680) and 278 (2680); $\nu_{\text{max}}/\text{cm}^{-1}$ 3497 (OH) and 3179 (OH); δ_{H} (CDCl₃) 5.27 (1H, s, 3-H), 5.51 (1H, s, 3-H), 5.91 (1H, s, 1-H), 6.79 (1H, dt, *J* 1.3 and 7.5, 5'-H), 6.89 (1H, dd, *J* 1.2 and 8.2, 3'-H), 6.98 (1H, dd, *J* 1.7 and 7.6, 6'-H), 7.18 (dt, 1H, *J* 1.7 and 7.7, 4'-H), 7.28–7.41 (5H, m, Ar-H) and 7.7 (1H, br s, OH); δ_{C} (CDCl₃) 76.3 (C-1), 115.5 (C-3), 117.2 (C-3'), 119.9 (C-5'), 124.7 (C-1'), 127.6 (C-2'' and -6''), 128.1 (C-4''), 128.5 (C-6'), 128.6 (C-3'' and -5''), 129.4 (C-4'), 138.7 (C-1''), 148.6 (C-2) and 155.8 (C-2'); *m/z* 226 (M⁺, 3%), 222 (8), 209 (16), 208 (M⁺ - 18, 100), 207 (93), 194 (10), 178 (26), 165 (10), 131 (30) and 104 (15) (Found: M⁺, 226.0996. C₁₅H₁₄O₂ requires *M*, 226.0993).

1-(2-Hydroxy-4-methoxyphenyl)-2-(4-hydroxyphenyl)prop-2-en-1-ol 3d

Flash chromatography eluent: *n*-hexane–acetone (1:1) Amorphous solid. λ_{max} (EtOH)/nm 206 ($\epsilon/\text{dm}^3 \text{mol}^{-1} \text{cm}^{-1}$ 41 700), 237 (12 600) and 260 (15 400); $\nu_{\text{max}}/\text{cm}^{-1}$ 3354 (OH); δ_{H} ([²H₆]–acetone) 3.68 (3H, s, OCH₃), 5.35 (1H, s, 3-H), 5.39 (1H, s, 3-H), 5.99 (1H, s, 1-H), 6.30 (1H, dd, *J* 2.4 and 8.4, 5'-H), 6.36 (1H, d, *J* 2.6, 3'-H), 6.73 (2H, d, *J* 8.6, 3''-H and 5''-H), 7.01 (1H, d, *J* 8.4, 6'-H), 7.30 (2H, d, *J* 8.8, 2''-H and 6''-H); δ_{C} ([²H₆]–acetone) 55.4 (OCH₃), 72.1 (C-1), 102.4 (C-3), 105.7 (C-3'), 111.5 (C-5'), 115.8 (C-3'' and -5''), 121.4 (C-1'), 128.9 (C-2'' and -6''), 129.9 (C-6'), 132.2 (C-1''), 151.1 (C-2), 157.6 (C-2'), 157.9 (C-4') and 161.1 (C-4''); *m/z* 272 (M⁺, 8%), 268 (6), 255 (18), 254 (100), 253 (65), 239 (26), 161 (9), 153 (21), 127 (12) and 121 (16) (Found: M⁺, 272.1047. C₁₆H₁₆O₄ requires *M*, 272.1048).

Acknowledgements

Funding to A. S. from the Foundation of Emil Aaltonen and to K. W. from the University of Helsinki is gratefully acknowledged. We thank Dr Jorma Matikainen for running the mass spectra.

References

- 1 M. A. Fitzgerald, P. J. M. Gunning and D. M. X. Donnelly, *J. Chem. Soc., Perkin Trans. 1*, 1976, 186.

- 2 B. C. B. Bezuidenhoudt, E. V. Brandt, D. G. Roux and P. H. van Rooyen, *J. Chem. Soc., Perkin Trans. 1*, 1980, 2179.
- 3 R. G. Cooke and I. D. Rae, *Aust. J. Chem.*, 1964, **17**, 379.
- 4 F. E. King, T. J. King and A. J. Warwick, *J. Chem. Soc.*, 1952, 1920.
- 5 B. C. B. Bezuidenhoudt, E. V. Brandt and D. G. Roux, *J. Chem. Soc., Perkin Trans. 1*, 1981, 263.
- 6 H. Adlercreutz, *Front. Gastrointest. Res.*, 1988, **14**, 165.
- 7 D. A. Shutt and R. I. Cox, *J. Endocrinol.*, 1972, **52**, 299.
- 8 T. J. Batterham, D. A. Shutt, N. K. Hart, A. W. H. Braden and H. J. Tweeddale, *Aust. J. Agric. Res.*, 1971, **22**, 131.
- 9 A. W. H. Braden, R. I. Thain and D. A. Shutt, *Aust. J. Agric. Res.*, 1971, **22**, 663.
- 10 H. Adlercreutz, P. I. Musey, T. Fotsis, C. Bannwart, K. Wähälä, T. Mäkelä, G. Brunow and T. Hase, *Clin. Chim. Acta*, 1986, **158**, 147.
- 11 C. Bannwart, H. Adlercreutz, T. Fotsis, K. Wähälä, T. Hase and G. Brunow, *Finn. Chem. Lett.*, 1984, **11**, 120.
- 12 H. Adlercreutz, T. Fotsis, C. Bannwart, K. Wähälä, T. Mäkelä, G. Brunow and T. Hase, *J. Steroid Biochem.*, 1986, **25**, 791.
- 13 H. Adlercreutz, T. Fotsis, C. Bannwart, K. Wähälä, G. Brunow and T. Hase, *Clin. Chim. Acta*, 1991, **199**, 263.
- 14 H. Adlercreutz, H. Markkanen and S. Watanabe, *Lancet*, 1993, **342**, 1209.
- 15 L. Coward, M. Kirk, N. Albin and S. Barnes, *Clin. Chim. Acta*, 1996, **247**, 121.
- 16 H. Adlercreutz, T. Fotsis, M. S. Kurzer, K. Wähälä, T. Mäkelä and T. Hase, *Anal. Biochem.*, 1995, **225**, 101.
- 17 M. S. Kurzer, J. W. Lampe, M. C. Martini and H. Adlercreutz, *Cancer Epidemiol. Biomarkers Prev.*, 1995, **4**, 353.
- 18 H. Adlercreutz, T. Fotsis, C. Bannwart, T. Mäkelä, K. Wähälä, G. Brunow and T. Hase, *Advances in Mass Spectroscopy – 85, Proceedings of the 10th International Mass Spectroscopy Conference*, ed. J. F. L. Todd, John Wiley, Chichester, 1986, p. 2179.
- 19 K. R. Price and G. R. Fenwick, *Food Add. Contam.*, 1985, **2**, 73.
- 20 G. E. Kelly, C. Nelson, M. A. Waring, G. E. Joannou and A. Y. Reeder, *Clin. Chim. Acta*, 1993, **223**, 9.
- 21 G. E. Joannou, G. E. Kelly, A. Y. Reeder, M. Waring and C. Nelson, *J. Steroid Biochem. Mol. Biol.*, 1995, **54**, 167.
- 22 S. Heinonen, K. Wähälä and H. Adlercreutz, *Anal. Biochem.*, 1999, in press.
- 23 V. N. Gupta and T. R. Seshadri, *Proc. Indian Acad. Sci., Sect. A*, 1956, **44**, 223.
- 24 A. C. Jain and P. Paliwar, *Indian J. Chem., Sect. B*, 1988, **27**, 985.
- 25 K. Wähälä, PhD Thesis, University of Helsinki, 1992.
- 26 J. W. Clark-Lewis and R. W. Jemison, *Aust. J. Chem.*, 1965, **18**, 1791.
- 27 C. D. Foxall and J. W. W. Morgan, *J. Chem. Soc.*, 1963, 5573.
- 28 V. Szabó and E. Antal, *Acta Chim. Acad. Sci. Hung.*, 1976, **90**, 381.
- 29 Á. Major, Z. Nagy and M. Nógrádi, *Acta Chim. Acad. Sci. Hung.*, 1980, **104**, 85.
- 30 M. Süsse, S. Johne and M. Hesse, *Helv. Chim. Acta*, 1992, **75**, 457.
- 31 K. Wähälä and T. Hase, *J. Chem. Soc., Perkin Trans. 1*, 1991, 3005.
- 32 K. Wähälä, T. Hase and H. Adlercreutz, *Proc. Soc. Exp. Biol. Med.*, 1995, **208**, 27.
- 33 K. Wähälä, A. Salakka and H. Adlercreutz, *Proc. Soc. Exp. Biol. Med.*, 1998, **217**, 293.
- 34 R. A. Micheli, A. N. Booth, A. L. Livingston and E. M. Bickoff, *J. Med. Pharm. Chem.*, 1962, **5**, 321.

Paper 9/04946K